# The Efficacy of Bacteriophage Therapy in Orthopaedic Field: A Systematic Review

# I Made Gilang Pinggan Kalimantara<sup>1</sup>, Ida Bagus Gede Darma Wibawa<sup>2</sup>

<sup>1</sup>Medical Doctor, Department of Orthopaedic & Traumatology, Mangusada Badung General Hospital Badung, Bali, Indonesia

<sup>2</sup> Orthopaedic and Traumatology Surgeon, Department of Orthopaedic & Traumatology, Mangusada Badung General Hospital Badung, Bali, Indonesia

Corresponding Author: I Made Gilang Pinggan Kalimantara

DOI: https://doi.org/10.52403/ijrr.20241248

## ABSTRACT

**INTRODUCTION:** Bacterial infections are the cause of high mortality rates in the world. Antibiotics and multidrug work well together to treat the patient. The pathogens are harder to control and develop antibiotics resistance. In orthopaedic field, infections like osteomyelitis, septic arthritis, or infections of the bone, joints, or implants (periprosthetic joint infections [PJI]) can be difficult to resolve both microbiologically and clinically. Bacterial viruses (bacteriophages) are one possible substitute. Due to their great host specificity, lack of adverse effects, and safety for eukaryotic cells. these viruses provide novel advantages, such as the safe treatment of illnesses.

**MATERIAL AND METHODS:** Three databases were used for the bibliographical search: Google Scholar, PubMed, and ScienceDirect for case report related to bacteriophage therapy and antibiotic resistance in orthopaedics, which were published between 2014 and 2024, which based on PRISMA guidelines qualified 10 articles for systematic review.

**RESULT:** In orthopaedic cases biofilm formation is widely recognized as the main obstacle to effective prevention and treatment. Biofilm production is intimately related to bacteria's capacity to create chronic illnesses and the challenge of overcoming them. The biofilm-disrupting properties of certain phages make them a promising option for managing deviceassociated infections. The capacity of phages to enter a biofilm and then cling to the surface of the host bacteria to infect and lyse it. The effectiveness of the reticuloendothelial system's clearance and possibly the production of phage-specific antibodies, which could result in phage inactivation, determine how long phages stay in systemic therapy.

**CONCLUSION:** Bacteriophages is a safe and effective treatment option, regardless of whether it is used alone or in conjunction with antibiotics and/or surgery. Bacteriophages is particularly well-suited for inclusion in multidimensional strategies to address infections due to its adaptability versatility. Rather than and replacing antibiotics. **Bacteriophages** should complement their effects to enhance infection management.

*Keywords:* Bacteriophages, Phage Therapy, Antibiotic Resistance, Orthopaedic, Biofilm

#### **INTRODUCTION**

Bacterial infections are the cause of high mortality rates in the world. Antibiotics and multidrug work well together to treat the patient. The pathogens are harder to control and develop antibiotics resistance.<sup>[1]</sup> Since bacterial pathogens react to the severe

selection pressures exerted on them, it is not surprising that antibiotic resistance is growing quickly as a result of overuse and misuse.<sup>[2]</sup> Unfortunately, the development of new antibiotics and new classes of drugs which act differently from existing ones is both expensive and takes a long time. As a result, some of our most potent medications losing their effectiveness.<sup>[3,4]</sup> are In Indonesia, antibiotic resistance phenomena like silent pandemic because of the high mortality rate, obtained data from The Ministry of Health Republic Of Indonesia that 1.2 million patient died because of antibiotic resistance.<sup>[5]</sup> Therefore, there is an urgent need for alternate antimicrobial techniques. Bacterial viruses (bacteriophages/phage therapy) are one substitute.<sup>[6,7]</sup> possible Given their and ease of manipulation, versatility bacteriophages could have an impact on biotechnology, research, and medicine.<sup>[8]</sup>

This review article's goal is to support the varied community of researchers, scientists, and biotechnologists who are employing phages to further and expand the biotechnology field.

In 1915 and 1917, Felix d'Herrelle and Twort described Frederick first bacteriophages, which are bacterial viruses that only infect bacteria. Phages are small viruses that range in size from 20 to 200 nm.<sup>[9]</sup> Due to their great host specificity, lack of adverse effects, and safety for eukaryotic cells, these viruses provide novel advantages, such as the safe treatment of illnesses.<sup>[2]</sup> This minimizes harm to the native microflora because they only reproduce when the bacteria that are causing the infection are present. Furthermore, it is uncommon for phages to share genetic material.<sup>[2,9]</sup>

When it comes to orthopaedic surgeons, infections like osteomyelitis, septic arthritis, or infections of the bone, joints, or implants (periprosthetic joint infections [PJI], fracture-related infections [FRI] involving plates, screws, or intramedullary nails) can be difficult to resolve both microbiologically and clinically. Currently available treatments, such as antibiotics and surgery, have a 10–20% failure rate. Infection rates after elective orthopaedic surgery range from 0.7% to 4.2%, but in trauma cases, they can be significantly higher. After surgery, infection rates for closed low-energy fractures range from around 1% to over 30% for complex open tibia fractures.<sup>[10]</sup>

Phage therapy (PT), has been used to treat infections.<sup>[11]</sup> Phage flexibility would enable us to use the antibodies against the bacteria that have been exhibited on the phage surface, whereas PT can be used alone to treat a bacterial infection by lysing the bacterial cell.

This study can be used as material for reconsider and rediscover PT. By being compatible with their hosts, PT reduces the likelihood of subsequent infections. The use of antibiotics targets both pathogens and the patients' normal flora, and it may result from secondary infections or superinfections. Although no negative side effects have been documented during or following phage administration, secondary infections, allergies, and bacterial resistance are the most frequent side effects of antibiotic treatment.<sup>[2,8,9]</sup>

The effectiveness statistics from research on human patients with bacterial infections of different kinds who received phage therapy are summarized in this systematic review.

## MATERIALS & METHODS Materials

Three databases were used for the bibliographical search: Google Scholar, PubMed, and ScienceDirect. The terms "bacteriophage therapy" and "antibiotic resistance in orthopaedics," which were published between 2014 and 2024, were used. Boolean operators have been employed to define the search as follows: "bacteriophage therapy" OR "phage therapy AND orthopaedic antibiotic resistance."

## **Study Selection and Data Collection**

Full-text publications and clinical case reports or case series that were published in

English throughout the last ten years (1 January 2014 to 31 December 2024) were included in the review. This study's primary goals were to describe the use of physical therapy (PT) in human patients who were infected with different bacteria, particularly in the orthopaedic field, so that it would be possible to determine if the treatment was effective or not. The inclusion and exclusion criteria were used to choose the relevant papers.

Exclusion criteria for this study include: lack of use of PT in multidrug-resistancebased studies: metagenomics in bacteriophages without subsequent application in human patients; PT in animals; phages in foods; patients not working in orthopaedics; publications published outside of the selected time frame; and bibliographic materials such as systematic reviews. reviews. posters. conferences, book sections. and perspectives. Studies that were found to be duplicates in the search results were excluded.

The selection of publication consider to the PRISMA statement (Preferred Reporting Items for Systematic Reviews and Meta-analyses).

#### **Data Extraction**

To verify for duplicate results across the three databases, the chosen citations and their titles were imported into Microsoft Office Excel. Records that were duplicates were removed.

## **Data Analysis**

Reviewing the abstracts and titles in order to assess the records. Everyone who didn't fit the requirements for inclusion was eliminated. The final record was included in this evaluation after the entire texts were examined to eliminate articles that did not fit the inclusion criteria.



Figure 1 PRISMA flowchart illustrating the search and selection of the articles

# RESULT

Ten out of the 1316 documents published between 2014 and 2024 that were retrieved for screening satisfied all eligibility requirements. Publications described experiences in the United States (n = 4), Belgium (n = 2), Germany (n = 1), the Netherlands (n = 1), Latvia (n = 1), and Israel (n = 1). The ten articles comprised five male patients and five female patients. Big three bacterial species, including Pseudomonas aeruginosa (n = 2), Klebsiella pneumoniae (n = 3), and Staphylococcus aureus (n = 3), were found to be resistant to many medications. Nine of these ten patients received the phage mixtures via multiple methods of delivery, whereas one patient received them intravenously.

## DISCUSSION

Based on the outcomes of ten patients treated in ten chosen studies, it is feasible to conclude that PT produced encouraging results for the treatment of infections caused by various bacterial species, especially those that are difficult to manage, such as infections caused by bacteria resistant to multiple antibiotics. The results of nearly all of these cases in this systematic review are improved. By successfully lowering the bacterial concentration, PT enhanced results and prevented fatal infections.

Furthermore, by removing bacterial toxins and lysates, the purification process and dilution of delivered phages would further improve their safety and prevent any negative effects or immunological reaction.<sup>[3]</sup>

Combination therapy can successfully treat and prevent or reduce the development of bacterial resistance in clinical settings by having a synergistic effect. To demonstrate how sublethal antibiotic doses could increase bacterial production of lytic phages, the phage antibiotic synergy (PAS) technique was employed. This is most likely caused by the low dosage of antibiotics preventing bacterial cell division and boosting biomass, which shortens the latent period and increases the phages' burst size, enabling them to swiftly eliminate the remaining bacterial cells.<sup>[3]</sup> Nine patients in this research got combination therapy, which primarily worked in concert to dramatically lower the concentration of germs. In addition, one patient who had physical therapy without the use of antibiotics reported good results. Since phages are believed to be a part of the healthy flora, their ability to act as probiotics and immunomodulators is neither surprising nor inconvenient.<sup>[3]</sup> Investigating the impact of phage alone without infection on the human immune system is crucial since this also calls into question whether the effectiveness of PT is dependent on its antibacterial and anti-inflammatory response.

The effectiveness of PT is not solely due to the use of a single bacteriophage or in antibiotics. conjunction with When considering bacteriophages as a therapy option, a number of things need to be taken into account. First, the patient's clinical condition may determine how long the treatment will last and how effective it is; the patient's immune system is crucial to the short-, medium-, and long-term success of physical therapy. The second is phage dose and administration methods. A precise assessment of the infection's kind and severity is necessary to determine the best administration method and phage dosage.<sup>[3]</sup> In fact, phages have a number of significant characteristics that play a role. The ability of phages to self-amplify is one of their advantages that sets them apart from traditional antibiotics and adds to their effectiveness. Second, certain phages have polysaccharide depolymerases on their tail which degrade structures, can the extracellular matrix of bacteria linked to biofilms and thus serve as an adjuvant to phage infection.<sup>[22]</sup>

Author	Country	Patient	Bacteria	Drugs	Infection	Effect of PT	PT Route	Combination
Randolph	US	63 YO	S. aureus	MRSA	Distal phalang	Quick recovery	inject to the	not combine
Fish et al <sup>[12]</sup>		F			osteomyelitis	without any	infection site	with other
						indication of	0.7cc once weekly	antibiotic
						bacteria in 14	for seven weeks	
						days		
Edison J.	US	62 YO	K	Daptomycin,	Total knee	observed	40 intravenous	combine with
Cano et al		М	pneumonia	penicillin,	arthoplasty	improvements in	doses of a single	minocycline
[15]			e	vancomycin,	Dextra	the right lower	phage	
				minocycline,		extremity's range		
				merononom		of motion,		
				meropenem		arythoma		
						edema and pain		
Ran Nir-Paz	Israel	42 YO	Δ	Piperacillin/tazobact	Both lower	The wound is	IV 1 ml of each	combine with
et al $\begin{bmatrix} 14 \end{bmatrix}$	151401	M	baumannii	am meropenem	extremity Grade	still closed and	phage $(5x107)$	intravenous
ot ui			and K.	uni, meropeneni	IIIA open	dry, and the	PFU/ml) IV tid	(IV)
			pneumonia		fractures (left	patient did not	110,111,11,400	meropenem (2
			e		bicondylar tibial	have their limb		gr tid) and
					plateau fracture	amputated.		colistin
					with	-		$(4.5 \times 106)$
					compartment			units/ bid),
					syndrome and			
					right distal			
					femoral fracture)			
Anaïs	Belgium	30 YO	Klebsiella	Ampicillin,	Femur Fracture	Improvement of	Locally to the	combine with
Eskenazi et		F	pneumonia	Amoxicillin-		the patient's	infection for 6	IV
al [15]			e	clavulanic acid,		wounds in	days	ceftazidime/av
				Piperacillin-		objective		ibactam (2
				tazobactam,		clinical,		g/0.5 g, q8h);
				Temocillin,		microbiological,		ugecycline
				Cefuroxillie, Ceftazidime		overall condition		(100  ing, 12h)
				Cefotaxime				y1211), moviflovacin
				Cefenime				(400 mg
				Meropenem				(400 mg,
				Ertapenem.				ciprofloxacin

								(400 mg, q8h)
Tamta Tkhilaishvili et al <sup>[16]</sup>	Germany	80 YO F	K. pneumonia e,P. stuartii, P. aeruginosa, S. epidermidi s, S. haemolytic us	Oxacillin, piperacillin, piperacillin- tazobactam, ceftazidime, avibactam, imipenem, aztreonam, cefepime, meropenem; CIP, ciprofloxacin; LEV, levofloxacin; GEN, gentamicin, tobramycin, amikacin, fosfomycin, colistin, doxycycline, rifampin, vancomycin, daptomycin, trimethoprim- sulfamethoxazole	Chronic osteomyelitis of the femur following a gunshot injury and recurrent right knee PJI	The patient's mobility was adequate, the soft tissue at the surgery site was unremarkable, and there was no pain in the right knee.	During surgery, a 100 ml loading dose of purified bacteriophage was delivered locally. Five milliliters of bacteriophage solution were then administered every eight hours.	combine with colistin, meropenem, and ceftazidime
Claudia Ramirez- Sanchez et al <sup>[17]</sup>	USA	61 YO F	Staphyloco ccus aureus	vancomycin, cefazolin	TKR Prosthetic Joint Infection	Numerous cultures of the patient's wounds and synovial fluid have come out negative for S. aureus.	Intra-articular and IV 12h/day 2weeks, intraoperative	combine with IV cefazoline
Brieuc Van Nieuwenhuy se et al <sup>[18]</sup>	Belgium	13 YO F	C. hathewayi, P. mirabilis, F. magna	Clindamycin, rifampin, flucloxacillin, ciprofloxacin, piperacillin- tazobactam, amoxicillin, ceftriaxone,	infection of an allograft of pelvic bone following surgery to remove Ewing's sarcoma	The patient has not experienced any further infectious episodes after two years.	intraoperative	combine with clindamycin, ciprofloxacin, rifampin

				ceftazidime.				
				vancomvcin.				
				metronidazole				
Ann-Sophie	Netherla	76 YO	E. faecalis	Teicoplanin,	infected total hip	Two years later,	PO combination	combine with
NEUTS et al	nds	М		amoxicillin,	arthroplasty	when we	for 19 days	amoxicillin,
[19]				amikacin,		examined him in		doxycycline
				sulfamethoxazole +		our outpatient		
				trimethoprim.		clinic, he had no		
				cefotaxime.		hip issues and no		
				Clarithormycin.		fresh cultures		
				Dalacin.		had been		
				Doxycycline.		collected.		
				Erythromycin.				
				ceftazidime				
				Gentamicin.				
				Rifampicin				
				azithromycin.				
				ceftriaxon.				
				cefuroxime				
Karlis	Latvia	21 YO	P.	Meropenem,	Osteomyelitis of	The patient's	intraoperative and	combine with
Racenis et al		М	aeruginosa	colistin, piperacillin-	the femur	wounds	IV 8h/dav+via	ceftadizim-
[20]				tazobactam,		remained dry	irrigation catheter	avibactam,
				linezolid, and		and closed six	for 7 days	linezolid
				fluconazole		months after		
						treatment ended.		
						while laboratory		
						inflammatory		
						markers stayed		
						steady within		
						typical levels.		
James B.	USA	72 YO	S. aureus	Vancomycin,	Prosthetic joint	was effective in	Daily IV phage	combine with
Doub et al		М		daptomycin,	infection	eliminating the	for 3 days	IV
[21]				doxycycline		patient's severe	-	daptomycin
						persistent		· ·
						infection.		

For orthopedic-related cases examined in this systematic review, improved outcomes were observed, including a successful result in a patient with recalcitrant S. aureus prosthetic joint infection (PJI) treated with prolonged PT alongside surgery and antibiotics. In this scenario, the ability of staphylococci to form adherent. multilayered biofilms on implanted medical devices posed a significant challenge to treatment, despite the isolate's antibiotic susceptibilities. The biofilm-disrupting properties of certain phages make them a promising option for managing deviceinfections.<sup>[17]</sup> associated А favorable outcome for an osteomyelitis patient was also shown in this case report. Antibiotics frequently exhibit poor and insufficient penetration to such infection sites, which complicates their use in the treatment of osteomyelitis in addition to the potential for antibiotic resistance.<sup>[20]</sup> Numerous studies on animals have demonstrated the beneficial effects of bacteriophages in the treatment of osteomyelitis.<sup>[23,24]</sup> Their erythema, induration, and local edema diminished, and they became more active.

In the orthopaedic field, biofilm formation is widely recognized as the main obstacle to effective prevention and treatment. Biofilm production is intimately related to bacteria's capacity to create chronic illnesses and the challenge of overcoming them. Bacteria flourish in biofilms because they are protected from the patient's immune system and antimicrobial therapies. The expression of resistance genes, decreased bacterial growth rates, and restricted antimicrobial penetration are some of the processes that lead to antibiotic tolerance in biofilms. By binding to particular receptors on the surface of the bacterial cell and introducing their genetic material, phages fight bacteria. These strain-specific sensors could be teichoic acids or proteins found on the bacterial cell wall. Once inside, phages have two options: either they stay latent within the cell (lysogenic cycle) or they use the bacterial metabolism to multiply and eventually lyse the host cell, releasing new phage particles (lytic cycle). Important phases for PT include infection, lysis, penetration, absorption, dispersion, and phage release.<sup>[25]</sup> For phage therapy to be applied locally, absorption and distribution are typically not significant. The effectiveness of the reticuloendothelial system's clearance and possibly the production of phage-specific antibodies, which could result in phage inactivation, determine how long phages stay in systemic therapy. The capacity of phages to enter a biofilm and then cling to the surface of the host bacteria to infect and lyse it is known as penetration. These processes are called adsorption, infection, and lysis.<sup>[22]</sup>



The light of the bacterial strains' previously stated pathogenicity and the low antibiotic absorption in bone tissue. In order to tackle isolates implicated in orthopaedic implantassociated infections, orthopaedic surgeons need possess new tactics for creating innovative therapeutic techniques. PT has long been shown to be a promising antibacterial strategy, mainly due to its high specificity and effectiveness in killing targeted pathogenic bacteria. In light of the rising problem of antibiotic resistance worldwide, PT appears to be a secure and successful method of countering the effects of bacterial resistance. However, there are not enough studies that thoroughly evaluate the safety and effectiveness of PT.

There are two countries where treatment with phages is routinely available in Europe: Georgia and Poland (Russia probably also uses PT, but much less information is available). More recently, the Wound Care Center in Lubbock, Texas used PT.<sup>[1]</sup>

## **CONCLUSION**

According to this systematic study, PT is a safe and effective treatment option, regardless of whether it is used alone or in conjunction with antibiotics and/or surgery. PT is particularly well-suited for inclusion in multidimensional strategies to address infections due to its adaptability and versatility. Rather than replacing antibiotics, PT should complement their effects to enhance infection management. To achieve this, emphasis should be placed on evaluating safety and efficacy, standardizing protocols, and identifying suitable host ranges.

Despite the challenges associated with PT, its adoption can lead to improved treatment outcomes. Conducting clinical trials is a crucial next step to confirm its effectiveness and determine its role in cases of therapeutic failure.

Declaration by Authors Acknowledgement: None Source of Funding: None **Conflict of Interest:** The authors declare no conflict of interest.

#### REFERENCES

- Golkar Z, Bagasra O, Gene Pace D. Bacteriophage therapy: A potential solution for the antibiotic resistance crisis. J Infect Dev Ctries. 2014;8(2):129–36.
- 2. Aranaga C, Pantoja LD, Martínez EA, et al. Phage Therapy in the Era of Multidrug Resistance in Bacteria: A Systematic Review. Int J Mol Sci. 2022;23(9):0–20.
- 3. Al-Ishaq RK, Skariah S, Büsselberg D. Bacteriophage treatment: Critical evaluation of its application on world health organization priority pathogens. Viruses. 2021;13(1).
- 4. Hatfull GF, Dedrick RM, Schooley RT. Phage Therapy for Antibiotic-Resistant Bacterial Infections. Annu Rev Med. 2022;73:197–211.
- 5. RI KK. wamenkes dante ajak atasi masalah resistensi antibiotik akibat mikroba 2022. [Internet]. p. https://www.kemkes.go.id/article/view/2208 2400003/. Available from: https://www.kemkes.go.id/article/view/2208 2400003/wamenkes-dante-ajak-atasimasalah-resistensi-antibiotik-akibatmikroba.html
- Jones JD, Varghese D, Pabary R, et al. The potential of bacteriophage therapy in the treatment of paediatric respiratory infections. Paediatr Respir Rev [Internet]. 2022;44:70–7. Available from: https://doi.org/10.1016/j.prrv.2022.02.001
- Varela-Ortiz DF, Barboza-Corona JE, González-Marrero J, et al. Antibiotic susceptibility of Staphylococcus aureus isolated from subclinical bovine mastitis cases and in vitro efficacy of bacteriophage. Vet Res Commun. 2018;42(3):243–50.
- Genevière J, McCallin S, Huttner A, et al. A systematic review of phage therapy applied to bone and joint infections: an analysis of success rates, treatment modalities and safety. EFORT Open Rev. 2021;6(12):1148–56.
- Barros J, Melo LDR, Poeta P, et al. Lytic bacteriophages against multidrug-resistant Staphylococcus aureus, Enterococcus faecalis and Escherichia coli isolates from orthopaedic implant-associated infections. Int J Antimicrob Agents [Internet].

2019;54(3):329–37. Available from: https://doi.org/10.1016/j.ijantimicag.2019.0 6.007

- Moriarty TF, Kuehl R, Coenye T, et al. Orthopaedic device-related infection: Current and future interventions for improved prevention and treatment. EFORT Open Rev. 2016;1(4):89–99.
- 11. Furfaro LL, Payne MS, Chang BJ. Bacteriophage Therapy: Clinical Trials and Regulatory Hurdles. Front Cell Infect Microbiol. 2018;8(October):1–7.
- Fish R, Kutter E, Bryan D, et al. Resolving digital staphylococcal osteomyelitis using bacteriophage - a case report. Antibiotics. 2018;7(4):1–6.
- 13. Cano EJ, Caflisch KM, Bollyky PL, et al. Phage Therapy for Limb-threatening Prosthetic Knee Klebsiella pneumoniae Infection: Case Report and in Vitro Characterization of Anti-biofilm Activity. Clin Infect Dis. 2021;73(1):E144–51.
- Nir-Paz R, Gelman D, Khouri A, et al. Successful Treatment of Antibioticresistant, Poly-microbial Bone Infection with Bacteriophages and Antibiotics Combination. Clin Infect Dis. 2019;69(11):2015–8.
- 15. Eskenazi A, Lood C, Wubbolts J, et al. Combination of pre-adapted bacteriophage therapy and antibiotics for treatment of fracture-related infection due to pandrugresistant Klebsiella pneumoniae. Nat Commun. 2022;13(1).
- 16. Tkhilaishvili T, Winkler T, Müller M, et al. Bacteriophages as Adjuvant to Antibiotics for the Treatment of Periprosthetic Joint Infection Caused by Multidrug-Resistant Pseudomonas aeruginosa. Antimicrob Agents Chemother. 2020;64(1):1–5.
- 17. Ramirez-Sanchez C, Gonzales F, Buckley M, et al. Successful treatment of staphylococcus aureus prosthetic joint infection with bacteriophage therapy. Viruses. 2021;13(6):1–10.
- 18. Van Nieuwenhuyse B, Galant C, Brichard B, et al. A case of in situ phage therapy against staphylococcus aureus in a bone allograft polymicrobial biofilm infection: Outcomes and phage-antibiotic interactions. Viruses. 2021;13(10):1–12.

- 19. Neuts AS, Berkhout HJ, Hartog A, et al. Bacteriophage therapy cures a recurrent Enterococcus faecalis infected total hip arthroplasty? A case report. Acta Orthop. 2021;92(6):678–80.
- Racenis K, Rezevska D, Madelane M, et al. Use of Phage Cocktail BFC 1.10 in Combination With Ceftazidime-Avibactam in the Treatment of Multidrug-Resistant Pseudomonas aeruginosa Femur Osteomyelitis—A Case Report. Front Med. 2022;9(April):1–9.
- 21. Doub JB, Ng VY, Johnson AJ, et al. Salvage bacteriophage therapy for a chronic MRSA prosthetic joint infection. Antibiotics. 2020;9(5):1–6.
- 22. Onsea J, Wagemans J, Pirnay JP, et al. Bacteri ophage therapy as a treatment strategy for orthopaedi c-devi ce-related i nfecti ons: Where do we stand? Eur Cells Mater. 2020;39:193–210.
- Kishor C, Mishra RR, Saraf SK, et al. Phage therapy of staphylococcal chronic osteomyelitis in experimental animal model. Indian J Med Res. 2016;143(JANUARY):87–94.
- 24. Cobb LH, Park JY, Swanson EA, et al. CRISPR-Cas9 modified bacteriophage for treatment of Staphylococcus aureus induced osteomyelitis and soft tissue infection. PLoS One [Internet]. 2019;14(11):1–17. Available from:

http://dx.doi.org/10.1371/journal.pone.0220 421

25. Bhargava K, Nath G, Bhargava A, et al. Phage therapeutics: from promises to practices and prospectives. Appl Microbiol Biotechnol [Internet]. 2021;105(24):9047– 67. Available from: https://doi.org/10.1007/s00253-021-11695-z

How to cite this article: I Made Gilang Pinggan Kalimantara, Ida Bagus Gede Darma Wibawa. The efficacy of bacteriophage therapy in orthopaedic field: a systematic review. *International Journal of Research and Review*. 2024; 11(12): 439-448. DOI: https://doi.org/10.52403/firm.20241248

https://doi.org/10.52403/ijrr.20241248

\*\*\*\*\*